

Preparation and Comprehensive Characterization of a Calcium Hydroxyapatite Reference Material

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Numerous biological and chemical studies involve the use of calcium hydroxyapatite (HA), $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$. In this study detailed physicochemical characterization of HA, prepared from an aqueous solution, was carried out employing different methods and techniques: chemical and thermal analyses, x-ray diffraction, infrared and Raman spectroscopies, scanning and transmission microscopies, and Brunauer, Emmett, and Teller (BET) surface-area method. The contents of calcium (Ca^{2+}), phosphate (PO_4^{3-}), hydroxide (OH^-), hydrogenphosphate (HPO_4^{2-}), water (H_2O), carbonate (CO_3^{2-}), and trace constituents, the Ca/P molar ratio, crystal size and morphology, surface area, unit-cell parameters, crystallinity, and solubility of this HA were determined. This highly pure, homogeneous, and highly crystalline HA is certified as a National Institute of Standards and Technology (NIST) standard reference material, SRM 2910.

Key words: chemical analysis; crystal size; crystallinity; hydroxyapatite; infrared; morphology; preparation; Raman; solubility; surface area; thermal analysis; unit-cell parameters; x-ray diffraction.

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1. Introduction

Calcium hydroxyapatite (HA), $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$, is an important inorganic material in biology and chemistry [1-3]. Biological apatites, which are the inorganic constituents of bone, tooth enamel and dentin, are typ-

ically very variable in their composition and morphology, and contain different impurities (Mg^{2+} , K^+ , Na^+ , CO_3^{2-} , HPO_4^{2-} , Cl^- , F^- , etc.) [1]. In general, these impure biological apatites are designated as calcium deficient or non-stoichiometric apatites.

Synthetic HAs are frequently used as reference materials in biomineralization and biomaterial studies. The composition, physicochemical properties, crystal size and morphology of synthetic apatites are extremely sensitive to preparative conditions. Common impurity phases in synthetic apatites prepared by precipitation from supersaturated aqueous solutions are calcium phosphate compounds such as amorphous calcium phosphates (ACP) with variable compositions of $\text{Ca}_3(\text{PO}_4)_{2-2x}(\text{HPO}_4)_{3x} \cdot n\text{H}_2\text{O}$, octacalcium phosphate (OCP), $\text{Ca}_8(\text{HPO}_4)_2(\text{PO}_4)_4 \cdot 5\text{H}_2\text{O}$, and calcium hydrogenphosphate dihydrate (DCPD), $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$. In addition, the incorporation of various ions as trace impurities (hydrogenphosphate, carbonate, silicate ions, etc.) is very difficult to prevent in any preparative procedure of HA [3].

For control and reference purposes, it is important to have available pure and stoichiometric HA, or nearly stoichiometric HA, characterized in detail with respect to its chemical composition and numerous other important properties. To meet this need, a large amount of highly pure, homogeneous and highly crystalline HA was synthesized by precipitation from aqueous solution of calcium hydroxide and phosphoric acid and then rigorously characterized by chemical and thermal analyses, infrared (IR) and Raman spectroscopies, powder x-ray diffraction (XRD), scanning and transmission microscopies, and surface area and solubility product [4] measurements. The chemical composition and other analyzed properties of this HA qualify it as a standard reference material (NIST, SRM 2910) [5] and it is hereafter denoted as HA-SRM.

Synthetic HA occurs in two structural forms, hexagonal and monoclinic, which have minor structural differences [2]. The hexagonal HA form is usually formed by precipitation from supersaturated solutions at 25 °C to 100 °C and the monoclinic form of HA is primarily formed by heating the hexagonal form at 850 °C in air and then cooling to room temperature [6]. The overall XRD patterns of hexagonal and monoclinic HA are almost identical; however the pattern of monoclinic HA has additional weak lines whose intensities are less than 1 % of the strongest hexagonal HA line [7]. The HA-SRM analyzed here is composed of the hexagonal form (mass fraction of about 75 %) and of the monoclinic form (mass fraction of about 25 %) as determined by normalized additional XRD measurements of the weak line of monoclinic HA at $2\theta = 36.28^\circ$ [6-8]. Only the hexagonal form, the major component in HA-SRM, is discussed in this paper. Preparation and characterization of the monoclinic form of HA and dif-

ferences between the hexagonal and monoclinic HA will be discussed in a separate paper [8].

2. Experimental Section

2.1 Preparation

Calcium hydroxyapatite-standard reference material (HA-SRM) was synthesized by solution reaction of calcium hydroxide and phosphoric acid in accordance with the preparation of McDowell et al. [9]. In brief, about 5 L of distilled water was boiled for 60 min in a 7.5 L Teflon-coated pot equipped with an electric stirring paddle, a reflux condenser with a CO_2 -absorbing NaOH trap to protect from atmospheric CO_2 , and ports for introducing titrant and nitrogen gas. Calcium oxide (prepared from calcium carbonate heated for 3 h at 1100 °C) was added to the water. Phosphoric acid (concentration 2 mol/L) was added to the calcium oxide/calcium hydroxide slurry at a rate of 0.3 mL/min to 0.6 mL/min and to a final Ca/P molar ratio of 1.67. The reacting mixture was boiled for 2 d. The precipitated solid phase was allowed to settle, the supernatant decanted, and an equal volume of boiled distilled water was added. This suspension was boiled for another 2 d. These washing and boiling procedures were repeated four times until the pH of the supernatant was ≈ 6 ; at pH 6, any possible traces of anhydrous dicalcium hydrogenphosphate (DCPA) are converted into HA. The precipitate, collected by filtration, was thoroughly washed with acetone, and then dried at 105 °C for 1 d. The yield was about 1 kg.

2.2 Characterization

The HA-SRM was characterized using different methods and techniques. Twenty randomly selected samples were analyzed for both calcium and total phosphorus content. Four samples were analyzed for the content of phosphorus in the form of hydrogenphosphate (HPO_4^{2-}). Fourteen samples were analyzed for water content. Twelve samples were analyzed for carbonate content. The contents of silicon and other trace constituents were determined in one sample. The specific surface area was determined on twelve samples. In addition, scanning and transmission electron microscopy (SEM and TEM), x-ray diffraction (XRD), and infrared (IR) and Raman spectroscopies were employed for detailed characterization.

2.3 Chemical Analyses

2.3.1 Calcium Content

Calcium was determined by atomic absorption spectroscopy with a Perkin-Elmer Model 603 spectrophotometer¹ using an air-acetylene flame and the 442.7 nm wavelength line. Standard calcium solutions used for calibration contained weighed amounts of calcium carbonate (NIST SRM 915, dried at 250 °C for 2 h) and LaCl_3 in the concentration of about 4.08 mmol/L (about 1000 ppm). The calcium solutions were placed in volumetric flasks (Class A) having volume of $500 \text{ mL} \pm 0.2 \text{ mL}$ (later assumed as a standard uncertainty). For experimental details see Refs. [4] and [10].

2.3.2 Phosphorus Content

Phosphorus was determined colorimetrically [11] as the phosphovanadomolybdate complex with a Cary Model 219 spectrometer using a wavelength of 420 nm. Standard phosphate solutions (flasks Class A with volume of $100 \text{ mL} \pm 0.08 \text{ mL}$) used for calibration contained weighed amounts of potassium dihydrogenphosphate (Baker Ultrex Reagent, dried at 105 °C for 2 h) and vanadomolybdate reagent. For experimental details see Refs. [4] and [10].

2.3.3 Hydrogenphosphate Content

The Gee and Deitz method [11] with some modifications [12] was used for determination of the content of phosphorus in the form of hydrogenphosphate (HPO_4^{2-}) in HA. The HA-SRM sample was heated at 550 °C in air for 24 h to convert the hydrogenphosphate into pyrophosphate ($\text{P}_2\text{O}_7^{4-}$). One portion (A) of this heated sample ($\approx 9 \text{ mg}$) was dissolved in 1 mol/L HClO_4 (in 100 mL volumetric flask) and heated in a boiling water bath for 3 h to hydrolyze the whole content of $\text{P}_2\text{O}_7^{4-}$ into phosphate ions (PO_4^{3-}). Another portion (B) of heated HA-SRM ($\approx 9 \text{ mg}$) was freshly dissolved at 25 °C just prior to phosphate analysis to minimize hydrolysis of $\text{P}_2\text{O}_7^{4-}$ to phosphates. The phosphorus concentrations were determined in both samples as described in Section 2.3.2. The difference in phosphorus contents between samples B and A corresponds to

the content of $\text{P}_2\text{O}_7^{4-}$ in the heated HA-SRM and to the content of HPO_4^{2-} in the unheated HA-SRM sample.

2.3.4 Water Content

The water content was determined from mass loss by three different procedures: (a) The thermogravimetric analysis (TGA) was performed on five samples all in the temperature range from 30 °C to 850 °C (rate 10 °C/min) in a nitrogen atmosphere. (b) Six powdered samples (mass 200 mg to 500 mg) were heated at 850 °C in air at $\approx 50 \%$ relative humidity for times ranging from 16 h to 20 h. The samples were weighed after cooling for 5 min in a desiccator at ambient conditions. (c) Three of the powdered samples were pressed into pellets and heated at 1000 °C in a steam atmosphere (100 kPa) for 10 h. These samples were weighed after cooling for 5 min in a desiccator.

2.3.5 Carbonate Content

The carbonate (CO_3^{2-}) content was determined by heating $\approx 5 \text{ g}$ of the HA-SRM sample at 1200 °C to liberate CO_2 that was collected in an absorption cell containing a lithium hydroxide solution. Carbonates in the absorption-cell were determined by automatic coulometric titration. These analyses were done by Galbraith Laboratories, Knoxville, TN.

2.3.6 Silicate and Trace Elements Content

The content of silicon and 63 other elements were analyzed by inductively coupled plasma mass spectroscopy (IPS-MS) by Galbraith Laboratories, Knoxville, TN.

2.4 Transmission and Scanning Electron Microscopy

Transmission electron micrographs were obtained from crystals placed directly onto formvar- and carbon-coated nickel grids, or from crystals that were suspended in solution by brief sonication in pure ethanol. In the latter case, ethanol-suspended crystals were allowed to settle onto the support film after which the ethanol was extracted from the edges of the grid with filter paper. Ultrastructural images of the crystals were recorded by transmission electron microscopy at an accelerating voltage of 80 kV using a JEOL JEM 2000FX-II. The samples for scanning electron microscopy were coated with gold and examined with a scanning electron microscope JEOL 5300.

¹ Certain commercial equipment, instruments or materials are identified in this paper to foster understanding. Such identification does not imply recommendation or endorsement by the National Institute of Standards and Technology, the National Institutes of Health, or the ADA Foundation nor does it imply that the materials or equipment identified are necessarily the best available for the purpose.

2.5 Surface Area

The surface area was determined by the triple-point BET (Brunauer, Emmett, Teller) method [13] with nitrogen as the adsorbate gas and helium as an inert non-adsorbable carrier. The mole fractions of nitrogen in N_2/He flowing mixtures were 0.1, 0.2 and 0.3. The sample mass was about 200 mg.

2.6 Infrared Spectroscopy

IR transmission and second derivative spectra were recorded with a Perkin-Elmer Model 621 spectrometer and with a Nicolet Magna 550 spectrometer, respectively, from the HA-SRM powder suspended in KBr pellets.

2.6.1 IR Transmission Spectra

IR transmission spectra from 4000 cm^{-1} to 300 cm^{-1} were recorded at 48°C (temperature in instrument light beam) with a Perkin-Elmer Model 621 spectrometer purged with dry CO_2 -free air. KBr sample pellets were run versus a blank KBr pellet in the reference beam to cancel KBr impurity bands, mainly H_2O bands. KBr pellets were prepared by mixing (not grinding) the pre ground HA-SRM (0.8 mg and 4.0 mg; particle cluster size $\leq 5\text{ }\mu\text{m}$ composed of crystal sizes of $0.1\text{ }\mu\text{m}$ to $0.5\text{ }\mu\text{m}$) with 400 mg of IR quality KBr (about $20\text{ }\mu\text{m}$ to $40\text{ }\mu\text{m}$ particle sizes). Grinding the sample and KBr together was avoided to reduce additional moisture adsorption from the ground and smaller KBr particles. The HA-SRM and KBr were mixed in a steel capsule on a mechanical shaker and then pressed in a 13 mm diameter evacuated die under a total force of 53,380 N (12 000 pound-force) for 30 s. One die face was machined nonparallel to the second die face, by about 1° . This nonparallel die face produced a wedge-shaped pellet, which reduced spectral interference fringes (especially important for second derivative spectra described below). Spectral slit widths were about 6 cm^{-1} for wavenumbers above 2000 cm^{-1} and 3 cm^{-1} to 5 cm^{-1} for wavenumbers below 2000 cm^{-1} . The wavenumber standard uncertainty, calibrated against standard indene bands [14], was 1 cm^{-1} for sharp bands and several cm^{-1} for broad bands.

The ion charges for infrared and Raman bands of different ions are normally omitted in the text.

2.6.2 IR Second Derivative Spectra

Second derivative spectra of absorbance spectra for the ν_3 and $\nu_4\text{ PO}_4$ bands were obtained in the ranges 1120 cm^{-1} to 1000 cm^{-1} and 670 cm^{-1} to 530 cm^{-1} with a Nicolet 550 Magna spectrometer purged with dry CO_2 -free air. The instrumental and data collection conditions were: deuterated triglycine sulfate detector at room temperature, KBr beam splitter, 1 cm^{-1} resolution, 1000 scans, 0.12 cm^{-1} data spacing, Happ-Genzel apodization, no smoothing of $\nu_3\text{ PO}_4$ absorbance spectrum, 25-point smoothing of $\nu_4\text{ PO}_4$ absorbance spectrum, and Nicolet Omnic software to obtain second derivative spectra of the absorbance spectra. High quality absorbance spectra without interference fringes and with low noise are required to obtain meaningful second derivative spectra. To help achieve this, the following were done: (1) to reduce interference fringes, wedge-shaped KBr pellets were prepared as described above (400 mg, 13 mm diameter with thickness increasing from 1.0 mm to about 1.2 mm across the pellet), (2) to eliminate the introduction of possible fringes in the background spectrum, the background for the sample was obtained from the empty pellet holder (no blank KBr pellet) in the spectrometer; and (3) to increase signal to noise, high sample concentrations and resultant high absorbance values of about 1.5 were used; the pellets contained 0.24 mg and 1.0 mg of HA-SRM for $\nu_3\text{ PO}_4$ and $\nu_4\text{ PO}_4$ spectra, respectively. KBr has no bands or impurity bands in the investigated regions. The second derivative wavenumber positions for the ν_3 and $\nu_4\text{ PO}_4$ bands were determined with a standard uncertainty of 0.1 cm^{-1} .

2.7 Raman Spectroscopy

Raman spectra were recorded with a Spex Model 1401 spectrometer in the 4000 cm^{-1} to 50 cm^{-1} region using the 488.0 nm wavelength excitation from an argon ion laser and a power of 320 mW measured at the sample. Spectra were obtained from about 4 mg of sample powder that was tamped in a cylindrical well (2.5 mm diameter, 1 mm deep) in the center of an aluminum disk 1.5 mm thick and 13 mm in diameter followed by pressing under a sufficient force of about 71,170 N (16,000 pound-force) for 5 s to reduce disk thickness, constrict the sample well and compact the sample. The exciting radiation, upward and vertical, was focused on the compacted sample in the disk tilted about 30° from the incoming radiation direction. Scattered radiation was collected at 90° to the incoming

beam direction and detected by a RCA C31034 photomultiplier cooled to -25°C .

The scattered radiation from the sample was passed through a 488.0 nm filter² placed ahead of the spectrometer entrance slit to reduce the intensity of the 488.0 nm exciting line that was reflected from the opaque sample. This filter markedly reduced the intensity of the 488.0 nm line (about $10^{-4}\%$ of original); this enabled obtaining spectra to within about 50 cm^{-1} of the exciting line and also eliminated spurious “grating ghost” bands.

The spectral slit width was 3.5 cm^{-1} . The wavenumber standard uncertainty was $\approx 0.5\text{ cm}^{-1}$, based on calibration using standard neon emission lines [15] from a neon lamp.

The baseline (BL) was obtained by reflecting the 488.0 nm line from a piece of rough surface platinum foil placed in the normal sample position. One spurious band was observed in the BL at 468 cm^{-1} .

2.8 X-Ray Diffraction

The x-ray diffraction (XRD) patterns of the powdered HA-SRM samples (about 150 mg in an aluminum holder) were obtained in the range of $3^{\circ} 2\theta$ to $70^{\circ} 2\theta$ with a Rigaku DMAX 2200 diffractometer operating at 40 kV and 40 mA, producing graphite-monochromatized $\text{CuK}\alpha$ radiation with wavelength $\lambda = 0.15405945\text{ nm}$, and at a scan speed of $0.030^{\circ} 2\theta/\text{min}$. The relative intensities were determined as diffraction line heights relative to the most intense line normalized to the intensity of 100, with the Materials Data, Inc., JADE 6.1 XRD Patterns Processing software (MDI JADE 6.1).

For determination of diffraction line positions (2θ -values), two samples were prepared by mixing HA-SRM with pre ground silicon (Silicon Powder $2\theta/d$ -Spacing Standard, NIST SRM 640b) that served as an internal standard to correct 2θ -values of HA-SRM. The samples contained mass fractions of 88 % HA-SRM and 12 % silicon. Two separate scans with the speed of $0.012^{\circ} 2\theta/\text{min}$ were obtained for each sample. For each scan, the position of each HA-SRM and silicon diffraction line was determined with MDI JADE 6.1 as the average of four measurements using pseudo-Voigt and Pearson-VII profile functions (two measurements for each profile function).

The HA-SRM unitcell (lattice) parameters were calculated with the Least Squares Unit Cell Refinement

and Indexing for Personal Computer (LSUCRIPC) program³; the input data were 2θ -values and corresponding indices (hkl) of the eight diffraction lines in the range from $39^{\circ} 2\theta$ to $54^{\circ} 2\theta$, which have relative intensities above 10, and do not overlap with other HA-SRM and silicon diffraction lines. For each HA-SRM sample, the unit-cell parameters were calculated from the average 2θ -values determined from the two separate scans. The final HA-SRM unit-cell parameters are the average of the data for the two samples.

Diffraction theory predicts that the diffraction lines of a XRD powder pattern will be very sharp for a crystalline material consisting of sufficiently large and strain-free crystallites [16]; therefore, the XRD line broadening (peak width) inversely correlates with crystal size and lattice perfection. The term “crystallinity” is commonly used to represent the crystallite size and lattice perfection. For determination of diffraction line angular width at its half-height, the lines having hkl indices (200), (002), (102), (210), (310) and (004) were recorded earlier with a vertically mounted Rigaku Denki diffractometer system operating at 40 kV and 25 mA, producing graphite-monochromatized $\text{CuK}\alpha$ radiation with wavelength $\lambda = 0.15405945\text{ nm}$ (time constant 10, scale 500 counts/s, scan speed $0.03125^{\circ} 2\theta/\text{min}$). The diffraction line angular width, B , at its half-height above background was measured with an optical magnifier and expressed in $^{\circ} 2\theta$. The angular width (B) was corrected for instrumental line broadening (b) caused by instrument imperfections [16]. The corrected value of the angular width (β) expressed in $^{\circ} 2\theta$, was calculated from Warren’s equation [16]

$$\beta = (B^2 - b^2)^{1/2}.$$

A stoichiometric, highly crystalline monoclinic hydroxapatite (hc-HA) prepared by solid-state thermal reaction [17] was used as a reference substance in determination of the value of b (the angular width at the half-height of hc-HA diffraction lines). The b -values for hc-HA diffraction lines were determined for the same six lines as for HA-SRM.

The reciprocal of the β value ($1/\beta$) correlates to the crystallite size/perfection [16].

² Holographic Notch Plus Filter, Kaiser Optical Systems, Inc., Ann Arbor, MI 48103.

³ Least Squares Unit Cell Refinement, NDSU version Fargo 90.10.13.em after Appleman and Evans (1973), implementation by Roy G. Garvey, Department of Chemistry, North Dakota State University, Fargo, ND 58105-5516.

2.9 Statistical Analysis

Uncertainties were assessed by the CIPM (International Committee for Weights and Measures) approach [18]. The uncertainty of a measurement result commonly consists of several components. An estimated standard deviation called a standard uncertainty, u_i , represents a component of uncertainty. A combined standard uncertainty, u_c , was computed by the method of propagation of uncertainties [18,19] and represents at the level of one standard deviation the combined effects of all standard uncertainties, u_i 's. According to the CIPM recommendation [18] the uncertainty of a measurement result is expressed with expanded uncertainty, U . Results in this paper, except as noted, are expressed as mean value $\pm U$, where $U = 2u_c$.

3. Results and Discussion

3.1 Chemical Composition

3.1.1 Calcium

The mass fraction of calcium in HA-SRM varied from 38.78 % to 39.49 % with a mean value of 39.15 % \pm 0.10 % (Table 1).

3.1.2 Phosphorus

The mass fraction of the total phosphorus content in HA-SRM varied from 18.111 % to 18.235 % with a mean value of 18.181 % \pm 0.037 %.

3.1.3 Ca/P Molar Ratio

From the mean values of Ca and P contents the calculated Ca/P molar ratio was 1.664 \pm 0.005. This value is in agreement with the ratio of 1.6649 \pm 0.0005 independently determined for this HA-SRM by thermal-product analysis [6,8]. The Ca/P ratio of 1.664 for this HA-SRM is about 0.2 % below the stoichiometric value of 1.6667.

3.1.4 Hydrogenphosphate and Phosphate

The mass fraction of phosphorus present in the form of hydrogenphosphate ions (HPO_4^{2-}) was 0.191 % \pm 0.010 % and accordingly, the mass fraction of HPO_4^{2-} was 0.592 % \pm 0.030 % (Table 1). The mass fraction of phosphorus present in the form of phosphate ions (PO_4^{3-}) is the difference between mass fractions of the total phosphorus content (18.181 % \pm 0.037 %) and of phosphorus present as HPO_4^{2-} (0.191 % \pm 0.010 %), giving the mass fraction of phosphorus present as PO_4^{3-} of 17.99 % \pm 0.05 %. From this value the calculated mass fraction of PO_4^{3-} was 55.16 % \pm 0.15 % (Table 1).

Table 1. Chemical composition of calcium hydroxyapatite standard reference material (HA-SRM) along with the calculated number and total relative charge of constituent ions per HA-SRM unit cell^a

Constituent	Mass fraction (%)	n^b	Number of constituent ions/HA-SRM unit cell ^c	Total relative charge of constituent ions/HA-SRM unit cell ^d
Ca^{2+}	39.15 \pm 0.10	20	9.985 \pm 0.026	+19.970 \pm 0.051
PO_4^{3-}	55.16 \pm 0.15	20	5.937 \pm 0.016	-17.811 \pm 0.048
HPO_4^{2-}	0.592 \pm 0.030	4	0.063 \pm 0.003	-0.126 \pm 0.006
H_2O	1.59 \pm 0.05	5	0.902 \pm 0.028	0
CO_3^{2-}	0.032 \pm 0.002	12	0.00545 \pm 0.00034	-0.0109 \pm 0.0007
SiO_3^{2-}	0.0406 ^e	1	0.00546	-0.0109
Trace elements ^f	0.0181 ^f	1	0.00595 ^f	+0.0144
OH^-	3.37 ^h \pm 0.12		2.026 ^h \pm 0.070	-2.026 ^g \pm 0.070
Sum	99.95 \pm 0.22			0

^a All results expressed as mean value $\pm U$, where U is expanded uncertainty.
^b Number of replicate measurements.
^c Number of constituent ions normalized to six phosphate groups (5.937 PO_4 + 0.063 HPO_4).
^d Calculated from relative electrical charge of the constituent ion time number of the constituent ions.
^e Calculated from silicon content in Table 2.
^f From Table 2.
^g Calculated to balance total charge to 0.
^h Derived from calculated relative charge of -2.026^g.

The contents of PO_4^{3-} and HPO_4^{2-} expressed as molar fractions of the total phosphate content were 98.95 % and 1.05 %, respectively.

3.1.5 Water

The total mass loss (expressed as the mass fraction) of samples heated continuously from 30 °C to 900 °C in a nitrogen atmosphere was $1.70 \% \pm 0.05 \%$. This mass loss is primarily attributed to water loss based on water band intensity changes in the IR spectrum of HA-SRM heated at 105 °C and 850 °C. The HA-SRM water content is the difference between the mass fractions of the total mass loss ($1.70 \% \pm 0.05 \%$) and the water loss derived from hydrogenphosphate pyrolysis into pyrophosphate and thermal reaction of calcium pyrophosphate and HA forming β -tricalcium phosphate; the calculated mass fraction of water derived from these thermal/chemical reactions was $0.111 \% \pm 0.006 \%$. Therefore, the mass fraction of water in HA-SRM was $1.59 \% \pm 0.05 \%$ or 0.902 H_2O molecule per HA-SRM unit cell (Table 1).

The TG-curve for HA-SRM (Fig. 1) is shown in the temperature range from 30 °C to 900 °C; on the left ordinate is mass fraction and on the right ordinate is the corresponding calculated number of water layers progressively removed from the HA-SRM surface. The number of water layers on the HA-SRM surface was calculated from the HA-SRM surface area of $18.3 \text{ m}^2/\text{g}$ (Section 3.2) and a cross-sectional area of 0.115 nm^2 for an adsorbed water molecule [20] on the HA surface; one monolayer of water corresponds to the mass fraction of 0.47 %. Rootare and Craig [20] have carried out detailed studies of vapor phase adsorption of water on HA. They found that the water monolayer that is in

contact with the HA surface (chemisorbed layer) was more strongly bound than the additional water layers (all physisorbed layers) that involved water/water contacts only. To completely remove the chemisorbed monolayer, heating at 300 °C in vacuum was required whereas the physisorbed layers could be removed at 20 °C in vacuum.

The TG-curve (Fig. 1) showed an initial mass loss (expressed as mass fraction) of $\approx 0.4 \%$ in the temperature range from 30 °C to 100 °C and a mass loss of $\approx 0.3 \%$ in the range from 100 °C to 250 °C. These two losses (mass fractions), giving a sum of $\approx 0.7 \%$, correspond to ≈ 1.5 layers mainly of physisorbed water although some chemisorbed water is also expected to be lost between 100 °C and 250 °C [20]. Between 250 °C and 360 °C, a loss of $\approx 0.55 \%$ was observed which corresponds to ≈ 1 layer of chemisorbed water. This temperature range, 250 °C to 360 °C, and mass loss equivalent to ≈ 1 water layer are consistent with data of Rootare and Craig [20] for the chemisorbed water layer. The mass fraction lost in the temperature range from 360 °C to 850 °C was $\approx 0.45 \%$. Of this $\approx 0.45 \%$, $\approx 0.11 \%$ corresponds to water loss from $\text{HPO}_4^{2-}/\text{P}_2\text{O}_7^{4-}/\text{HA}/\beta\text{-TCP}$ reactions, $\approx 0.02 \%$ corresponds to loss from CO_3^{2-} decomposition on heating to 850 °C and the remainder of $\approx 0.32 \%$ corresponds to ≈ 0.7 layer of water that is more strongly held by the crystals than the chemisorbed layer.

From these TG-data it appeared that the total number of water layers at the surface of the HA-SRM crystals was ≈ 2.5 ; ≈ 1.5 layers correspond to physisorbed water and ≈ 1 layer to chemisorbed water. The location of the more strongly-held water, equivalent to ≈ 0.7 layer or about one water molecule per 5.6 HA-SRM unit cells is uncertain. It may be “structural” water or water trapped within crystals.

The mass fraction of water in HA-SRM determined from mass loss of powdered HA-SRM samples heated in air at 850 °C for 16 h to 20 h, then cooled in a desiccator and weighed in the laboratory atmosphere (50 % relative humidity) at ambient temperature was $1.430 \% \pm 0.034 \%$, whereas the mass fraction of water in HA-SRM determined in samples pressed into pellets and heated in a steam atmosphere at 1000 °C for 10 h and then cooled and weighed as above was $1.564 \% \pm 0.028 \%$. In both cases the HA-SRM water content was lower than in the samples heated and weighed in the nitrogen atmosphere because of fast readsorption of surface water during cooling and weighing in the air atmosphere at ambient temperature.

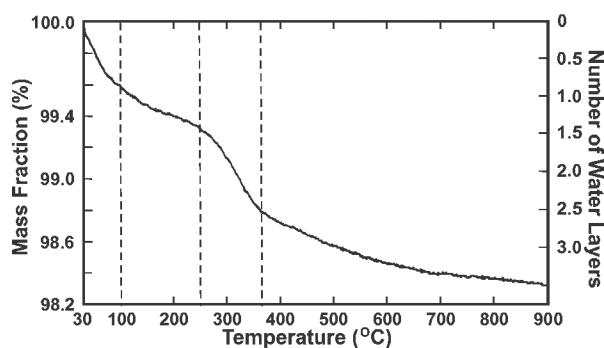


Fig. 1. TG-curve for HA-SRM in the temperature range from 30 °C to 900 °C in a nitrogen atmosphere. The left ordinate denotes the mass fraction and the right ordinate gives the corresponding calculated number of water layers progressively removed from the HA-SRM surface.

3.1.6 Carbonate

Carbonate ions are a common impurity in HA. The mass fraction of carbonate found in HA-SRM was in the range from 0.029 % to 0.033 % with the mean value of 0.032 % \pm 0.002 % (Table 1). This carbonate content corresponds to 0.00545 CO₃²⁻ ion per HA-SRM unit cell (Table 1) or to one CO₃²⁻ ion per 183 HA-SRM unit cells.

3.1.7 Silicate

The mass fraction of silicon of 0.015 % (Table 2) expressed as mass fraction of silicate ions, SiO₃²⁻, was 0.0406 % (Table 1). This content corresponds to 0.00546 SiO₃²⁻ ion per HA-SRM unit cell or to one SiO₃²⁻ ion per 183 HA-SRM unit cells. The source of the silicon impurity was most plausibly the boro-silicate glass apparatus used in preparation of HA-SRM.

3.1.8 Trace Constituents

Trace constituents with mass fractions above 0.0005 % (>5 ppm) in HA-SRM are listed in Table 2 and summarized in Table 1. Approximately 0.001 atom each of Al, B, Mg, Na and Sr occurs per HA-SRM unit cell (Table 2), which corresponds to approximately one of each atom per 1000 unit cells. The sum of trace constituent atoms of 0.00595 per HA-SRM unit cell (Table 1) corresponds to one trace constituent atom per 168 HA-SRM unit cells.

Table 2. Contents of trace constituents^a and silicon in HA-SRM

Trace constituent	Mass fraction (%)	Number of ions/HA-SRM unit cell ^b
Al ³⁺	0.0029	0.00110
Ba ²⁺	0.0024	0.00018
B ³⁺	0.0015	0.00142
Mg ²⁺	0.0029	0.00122
Na ⁺	0.0031	0.00138
Sr ²⁺	0.0044	0.00051
Zn ²⁺	0.0009	0.00014
Sum	0.0181	0.00595
Si	0.0150	0.00546

^a Trace constituents having mass fraction >0.0005 % are included.

^b Calculated number of ions per unit-cell.

3.1.9 Hydroxide

In Table 1 are listed the contents of analyzed HA-SRM constituents: calcium, phosphate, hydrogen-phosphate, water, carbonate, silicate and sum of trace constituents. From these contents the number of constituents per HA-SRM unit cell was calculated by normalizing the total number of phosphate groups (PO₄³⁻ and HPO₄²⁻) to six, 5.937 PO₄³⁻ and 0.063 HPO₄²⁻. The relative charge attributed to the total number of hydroxide ions (OH⁻) per unit cell was calculated from the difference between positive and negative relative charges of all unit-cell constituents; a mean value of -2.026 for OH⁻ ions balanced the total charge to zero. This calculated number of 2.026 \pm 0.070 of OH⁻ ions per HA-SRM unit cell corresponds to the mass fraction of 3.37 % \pm 0.12 % of OH⁻ in HA-SRM (Table 1).

3.1.10 Sum of Mass Fractions

The total sum of mass fractions of all constituents was 99.95 % \pm 0.22 % (Table 1); this shows high accuracy of the chemical analyses.

3.2 Crystal Morphology and Specific Surface Area

Transmission and scanning electron micrographs of the HA-SRM crystals are shown in Fig. 2. Generally, the crystals appear to have a cylindrical shape with heights of \approx 0.1 μ m to 0.3 μ m and diameters of \approx 0.05 μ m to 0.15 μ m. The specific surface area determined by BET was 17.7 m²/g to 19.1 m²/g with an average value of 18.3 m²/g \pm 0.3 m²/g. This specific surface area for the HA-SRM crystals compares well with the value of 16.6 m²/g calculated by assuming an average cylindrical particle with height of 0.2 μ m and diameter of 0.1 μ m.

The HA sample of McDowell et al. prepared by precipitation from solutions had a specific surface area of 16.7 m²/g determined by BET [9]. This value of 16.7 m²/g is in agreement with the above value of 18.3 m²/g for HA-SRM. These data indicate the reproducibility of crystal sizes and surface area of HAs prepared by the same method.

3.3 Infrared Spectra

3.3.1 IR Transmittance Spectra

IR transmittance spectra of HA-SRM at two different concentrations (0.8 mg and 4.0 mg HA-SRM per

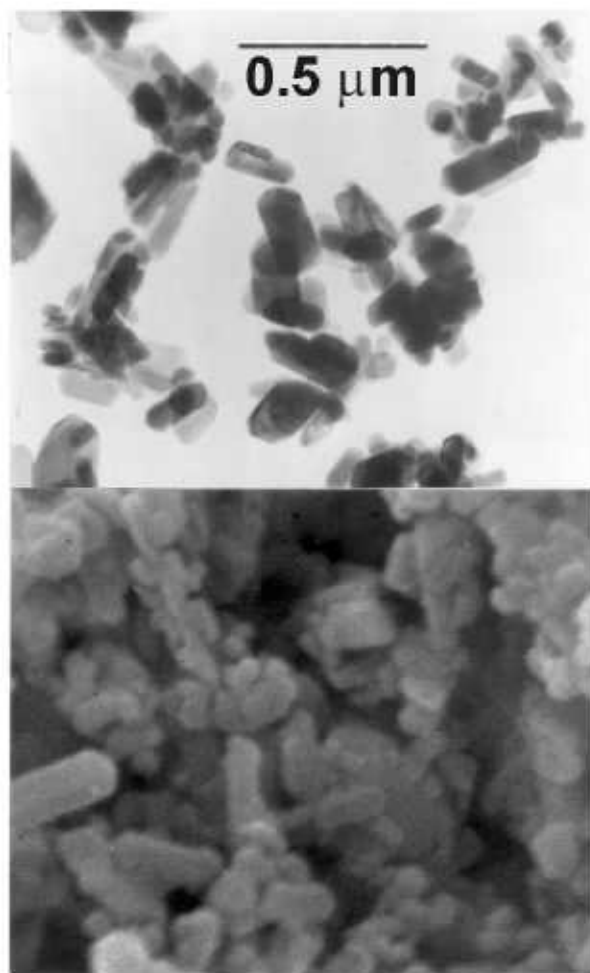


Fig. 2. Transmission electron micrograph (top) and scanning electron micrograph (bottom) of the HA-SRM crystals. Both micrographs have the same magnification and the bar length in the top micrograph is 0.5 μm .

400 mg KBr) are shown in the 4000 cm^{-1} to 300 cm^{-1} range in Fig. 3. The spectra show the bands of HA along with additional bands that are ascribed to impurity ions (CO_3^{2-} , HPO_4^{2-} , and silicate ions), and associated H_2O .

Bands of HA [21]: (a) The bands at 3572 cm^{-1} , 631 cm^{-1} , and 342 cm^{-1} arise from stretching, librational, and translational modes, respectively, of OH^- ions. (b) The 1090 cm^{-1} and about 1040 cm^{-1} bands arise from $\nu_3\text{ PO}_4$, the 962 cm^{-1} band arises from $\nu_1\text{ PO}_4$, the 601 cm^{-1} and 574 cm^{-1} bands arise from $\nu_4\text{ PO}_4$, and the 472 cm^{-1} band arises from $\nu_2\text{ PO}_4$. (c) The group of weak intensity bands in the 2200 cm^{-1} to 1950 cm^{-1} region derives from overtones and combinations of the ν_3 and $\nu_1\text{ PO}_4$ modes. The sharpness of bands, especial-

ly sharpness of the 631 cm^{-1} , 601 cm^{-1} , and 574 cm^{-1} bands, indicate a well-crystallized HA.

Bands of CO_3^{2-} impurity ions: The weak intensity bands at about 1410 cm^{-1} and 1450 cm^{-1} in the spectrum of HA-SRM at high concentration (4.0 mg of HA-SRM per 400 mg KBr) are attributed to components of the ν_3 mode of a trace amount of CO_3^{2-} . The mass fraction of CO_3^{2-} in HA-SRM determined by chemical analysis, Sec. 3.1.6, was 0.032 %. The areas and intensities of these CO_3 bands correspond to mass fraction of about 0.03 % CO_3^{2-} by comparison to CO_3 bands of other HA samples [6] containing chemically analyzed CO_3^{2-} mass fractions of about 0.3 %; this band intensity agreement for this low CO_3^{2-} content helps identify these weak intensity bands as CO_3 bands. Bands of other CO_3 modes, ν_4 and ν_1 , were not detected because of their weak intensities and the $\nu_2\text{ CO}_3$ band at about 872 cm^{-1} , with intensity about one fifth that of $\nu_3\text{ CO}_3$, is obscured by the HPO_4 band at 875 cm^{-1} . The CO_3 bands at 1410 cm^{-1} and 1450 cm^{-1} derive from CO_3^{2-} (designated the “B-type” carbonate) that replace PO_4^{3-} ions in the HA lattice [22] (and references therein). Bands at 1455 cm^{-1} and about 1540 cm^{-1} , which derive from CO_3^{2-} (designated the “A-type” carbonate) that replace OH^- ions in the HA lattice [23], were not detected. The mass fraction of 0.032 % chemically determined CO_3^{2-} , corresponds to one CO_3^{2-} ion per 1101 total phosphate ions (PO_4^{3-} and HPO_4^{2-}).

Bands of HPO_4^{2-} impurity ions: The band at 875 cm^{-1} is attributed to arise from HPO_4^{2-} ions for several reasons [24,25]. Chemical analysis shows that HA-SRM contains 1.05 HPO_4^{2-} ions per 98.95 PO_4^{3-} ions (Sec. 3.1.4, Table 1) or molar fraction of 1.05 % HPO_4^{2-} with respect to the total P content. The isolated HPO_4^{2-} ion has 9 predicted infrared active internal modes for its highest symmetry point group, C_{3v} , and 12 predicted infrared active modes for its lowest symmetry point group, C_1 . At this very low molar fraction of 1.05 % HPO_4^{2-} , of the 9 to 12 possible bands, only the 875 cm^{-1} band is clearly detectable; the other HPO_4 bands are obscured by the PO_4 bands of HA and, in addition, the ($-\text{O}-\text{H}$) bands of the HOPO_3^{2-} ions are broad and weak in intensity. The normalized intensity and area of the 875 cm^{-1} band correlates with HPO_4^{2-} content determined by chemical analysis. A HA sample containing a HPO_4^{2-} molar fraction of 2.34 % by chemical analysis [3,6] had a 875 cm^{-1} normalized band area 2.1 times that of the HA-SRM that contained HPO_4^{2-} molar fraction of 1.05 % determined by chemical analysis. In addition, this 875 cm^{-1} HPO_4 band was, as expected, missing in spectra of HA-SRM that had been heated at

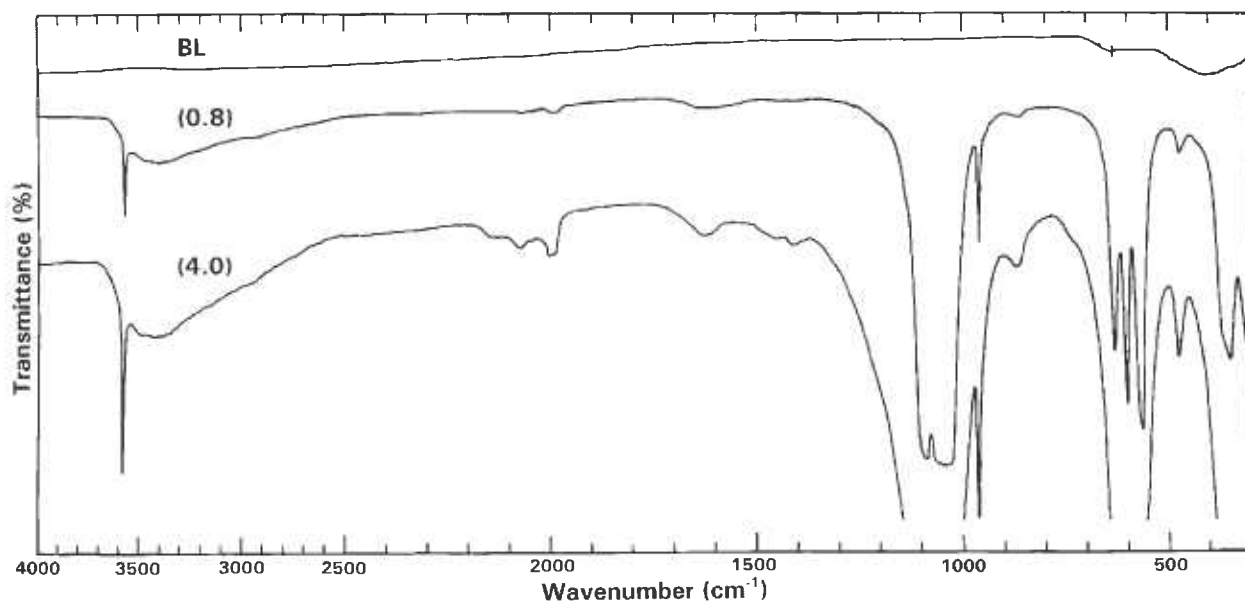


Fig. 3. Infrared percent transmittance spectra of HA-SRM from concentrations of 0.8 mg and 4.0 mg of HA-SRM/400 mg of KBr in the 4000 cm^{-1} to 300 cm^{-1} region. BL denotes the KBr pellet baseline.

550 $^{\circ}\text{C}$ because of condensation of HPO_4^{2-} ions to form $\text{P}_2\text{O}_7^{4-}$ ions and H_2O .

Bands of silicate impurity ions: The mass fraction of Si in HA-SRM determined by chemical analyses was 0.015 % (Sec. 3.1.7); the mass fraction calculated as the SiO_3^{2-} was 0.0406 % (Table 1). Previous work [6] on other HAs prepared by precipitation in glass apparatus from solution at 100 $^{\circ}\text{C}$ and high pH produced HAs that contained Si mass fraction of about 0.1 % to 0.3 % determined by chemical analyses. IR spectra of these HAs had weak bands, not deriving from HA, at 890 cm^{-1} , $\approx 830 \text{ cm}^{-1}$, $\approx 750 \text{ cm}^{-1}$ and $\approx 500 \text{ cm}^{-1}$ and a Raman band at 890 cm^{-1} whose intensities correlated with silicon content. Consequently, these bands were attributed to silicate ions, and their most probable source was the glass apparatus. The type of silicate ion SiO_3^{2-} (chain or ring structures), $\text{Si}_2\text{O}_7^{6-}$, or SiO_4^{4-} in these HAs was not identified with certainty by IR or Raman methods primarily because of the low silicate contents and resultant weak band intensities along with interference from the strong HA bands. Nevertheless, the combined IR and Raman data and additional thermal data suggested that $(\text{SiO}_3^{2-})_3 = \text{Si}_3\text{O}_9^{6-}$ ring and $\text{Si}_2\text{O}_7^{6-}$ ions may be present and SiO_4^{4-} and acidic silicates less probable. The high concentration spectrum of HA-SRM in Fig. 3 has very weak bands at 890 cm^{-1} and $\approx 750 \text{ cm}^{-1}$; these two bands are better discerned in the high concentration spectrum of the heated HA-SRM that will be shown in the paper on monoclinic HA [8].

These 890 cm^{-1} and 750 cm^{-1} bands are attributed to silicate ions and are assumed to be SiO_3^{2-} ions.

Bands of H_2O molecules: The broad band from about 3700 cm^{-1} to 2500 cm^{-1} derives from the ν_3 and ν_1 stretching modes of hydrogen-bonded H_2O molecules, and the band at 1630 cm^{-1} derives from the ν_2 bending mode of the H_2O molecules. The thermogravimetric data in Table 1 show a mean mass loss (expressed as mass fraction) of 1.59 % on heating HA-SRM that is primarily attributed to loss of adsorbed water. In the IR spectra of HA-SRM after heating at 850 $^{\circ}\text{C}$ [8], the above water bands are, as expected, missing; this indirectly identifies H_2O as the principal component lost on heating.

3.3.2 IR Second Derivative Spectra

IR second derivative spectra of the ν_3 and ν_4 PO_4 bands are shown in Fig. 4 and Fig. 5, respectively, and the second derivative band positions are given in Table 3. Second derivative spectra of the ν_1 and ν_2 PO_4 bands are not shown. Only one ν_1 PO_4 band was detected at 962.9 cm^{-1} in second derivative spectra and the instrument detector response, progressively lower in the 500 cm^{-1} to 400 cm^{-1} region along with the weak ν_2 PO_4 band intensity, precluded obtaining well-resolved second derivative spectra of the ν_2 PO_4 band although the bands occur at about 474 cm^{-1} and 462 cm^{-1} . Under 1 cm^{-1} resolution, eleven ν_3 PO_4 bands were resolved

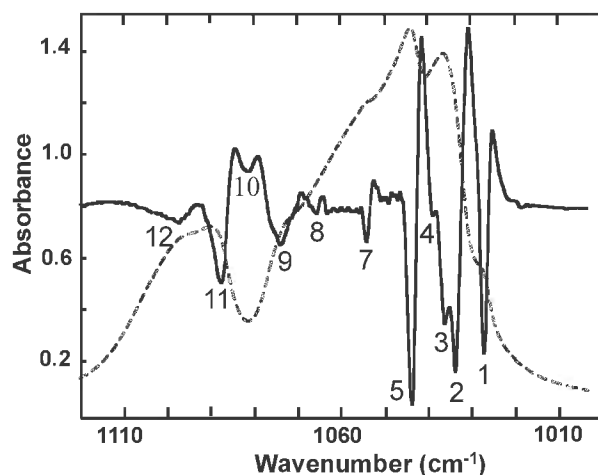


Fig. 4. Infrared absorbance spectrum of the ν_3 PO_4 mode of HA-SRM (dashed line) and second derivative of the absorbance spectrum (solid line). The second derivative spectrum ordinate scale, not shown, is arbitrary. The second derivative spectrum was adjusted to full ordinate range and the minima denoted by numbers identify band positions in the absorbance spectrum.

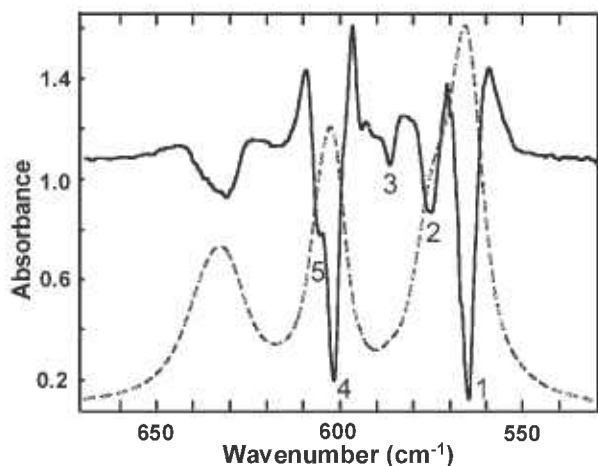


Fig. 5. Infrared absorbance spectrum of the ν_4 PO_4 mode of HA-SRM (dashed line) and second derivative of the absorbance spectrum (solid line). The absorbance band at 633 cm^{-1} and second derivative band at 633 cm^{-1} derive from the OH^- librational mode. The description of the second derivative spectrum is the same as that given in Fig. 4.

(Fig. 4). Two of these bands, numbered 3 and 4 in Fig. 4 and in Table 3, are attributed to arise from the mass fraction of about 25 % of monoclinic HA; these bands will be discussed in the paper on monoclinic HA [8]. Thus, nine bands were detected for the ν_3 PO_4 mode of this hexagonal HA-SRM. In Fig. 5, five second derivative ν_4 PO_4 bands were detected; the absorbance band and second derivative band at 633 cm^{-1} derive from the OH^- librational mode.

Table 3. IR wavenumber positions of ν_3 and ν_4 PO_4 bands of HA-SRM obtained from second derivative spectra

Band number ^a	PO_4 bands (cm^{-1})	
	ν_3	ν_4
1	1027.0	565.1
2	1033.7	575.3
3	1036.0 ^b	586.4
4	1038.7 ^b	601.8
5	1043.6	605.4
6		
7	1054.0	
8	1065.4	
9	1073.9	
10	1081.3	
11	1087.7	
12	1097.5	

^a Refer to Figs. 4 and 5.

^b These two bands are attributed to arise from monoclinic HA (mass fraction $\approx 25\%$).

3.4 Raman Spectra

Raman spectra of HA-SRM in the range from 4000 cm^{-1} to 50 cm^{-1} recorded with relative intensities of 1 and 10 in the range below 1200 cm^{-1} and with relative intensity of 3.3 in the range above 1200 cm^{-1} are shown in Fig. 6. The spectra have the bands of hexagonal HA and two additional bands that arise from HPO_4^{2-} impurity ions. Under the spectral resolution used (spectral slit width of 3.5 cm^{-1}), no bands of monoclinic HA are resolved.

Bands of HA [26-29]: (a) The 3573 cm^{-1} and 329 cm^{-1} bands arise from stretching and translational modes of the OH^- ions, respectively; the OH^- librational bands expected in the 630 cm^{-1} region are not clearly detected although two bands are predicted by C_6 factor group symmetry analysis [21]. (b) The 1076 cm^{-1} , 1052 cm^{-1} (shoulder, sh), 1047 cm^{-1} , 1040 cm^{-1} (sh), and 1028.5 cm^{-1} bands arise from ν_3 PO_4 , the very strong 962 cm^{-1} band arises from ν_1 PO_4 , the 614 cm^{-1} , 607 cm^{-1} , 590 cm^{-1} , and 579 cm^{-1} bands arise from ν_4 PO_4 , and the 447 cm^{-1} and 431 cm^{-1} bands arise from ν_2 PO_4 . (c) The group of weak intensity bands in the 329 cm^{-1} to 50 cm^{-1} region derives from translations of the Ca^{2+} , PO_4^{3-} , and OH^- ions and librations of the PO_4^{3-} ions. The 329 cm^{-1} , 305 cm^{-1} , and 270 cm^{-1} bands have been assigned to vibrations of the $2[(\text{Ca}_{11})_3(\text{OH})]$ sublattice of hexagonal HA, and the band at 285 cm^{-1} primarily to libratory phosphate motions [28,30].

Bands of CO_3^{2-} impurity ions: The strongest intensity CO_3 band, ν_1 , for the B-type CO_3^{2-} impurity occurs at 1070 cm^{-1} ; this band is obscured by the strong intensi-

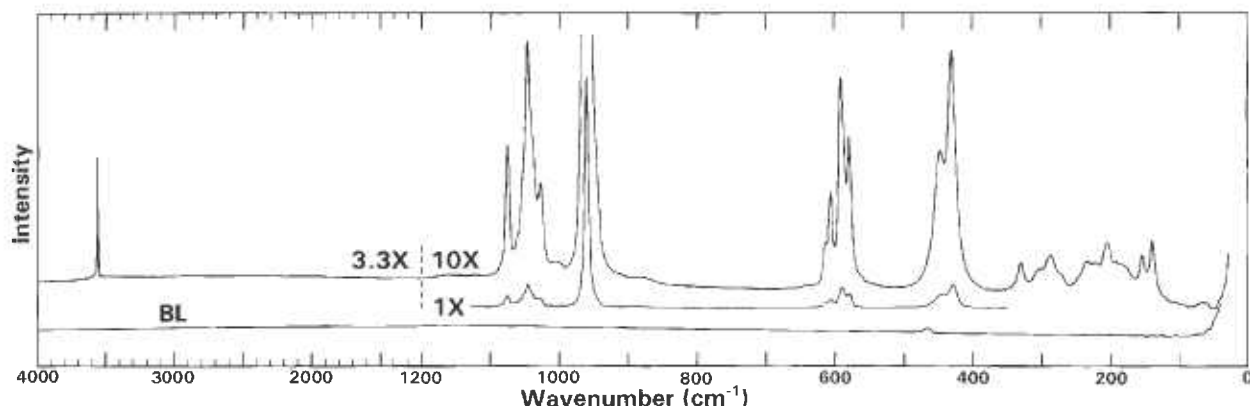


Fig. 6. Raman spectra of HA-SRM from 4000 cm^{-1} to 50 cm^{-1} recorded at relative intensities of 1 and 10 below 1200 cm^{-1} and 3.3 above 1200 cm^{-1} . BL denotes the baseline.

ty PO_4 band at 1076 cm^{-1} . The other CO_3 modes ν_3 , ν_4 , and ν_2 (ν_2 is expected to be Raman active because of low symmetry of CO_3^{2-} ion) have band positions not obscured by the PO_4 bands, but they have weak intensities and were not detected. The ν_1 CO_3 band for A-type CO_3^{2-} , unobscured by PO_4 bands, occurs at 1106 cm^{-1} [31], and is useful for detecting the A-type CO_3^{2-} . However, this band was absent; this was expected because the IR spectra did not have bands for the A-type CO_3^{2-} .

Bands of HPO_4^{2-} impurity ions: The weak band at 1005 cm^{-1} is assigned to symmetric stretching of the HPO_4^{2-} ions and the weak band at 880 cm^{-1} to [P-(OH)] stretching of the HPO_4^{2-} ions [6,32]. Similarly as in IR spectra, these two Raman bands increase in intensity with an increase in HPO_4^{2-} content, and they are missing in spectra of HA-SRM that had been heated at 550 $^\circ\text{C}$ because of thermal conversion of HPO_4^{2-} ions to $\text{P}_2\text{O}_7^{4-}$ ions. This independent detection of HPO_4^{2-} ions in Raman spectra confirms the IR data on HPO_4^{2-} ions.

Bands of silicate impurity ions: Bands of the trace silicate impurity, probably present as SiO_3^{2-} or $\text{Si}_2\text{O}_7^{6-}$ ions, were not detected because of the low silicate mass fractions (about 0.04 % as SiO_3^{2-} or $\text{Si}_2\text{O}_7^{6-}$ ions). The mass fraction of 0.2 % of silicate impurity (as $\text{Si}_2\text{O}_7^{6-}$) was detectable in other HA preparations by the 890 cm^{-1} band arising from $\text{Si}_2\text{O}_7^{6-}$ ions.

Bands of H_2O molecules: Water vibrational modes give rise to weak intensity stretching and bending bands in Raman spectra. The water component in HA-SRM (mass fraction of 1.59 %) causes IR bands at 3700 cm^{-1} to 2500 cm^{-1} and 1630 cm^{-1} ; these water bands, expected at about the same wavenumbers in Raman spectra, were not observed in Raman spectra under the spectral intensity expansion used in Fig. 6.

3.5 Combined Infrared and Raman Data

A rigorous comparison of the number and coincidences of the IR and Raman bands cannot be made with the present data because equivalent high-resolution second derivative Raman spectra were not obtained for HA-SRM. Although additional Raman bands may be detected, comparisons of the predicted and observed current data are meaningful and are given in Table 4. IR and Raman bands that have wavenumber positions within 2 cm^{-1} were considered coincident.

The number and coincidence or noncoincidence of the infrared and Raman active bands predicted according to factor group analysis for the ν_1 , ν_2 , ν_3 , and ν_4 PO_4 modes of HA having hexagonal structures ($\text{P6}_3/\text{m}$, C_{6h}) and (P6_3 , C_6) [21] are given in Table 4 along with the observed number and coincidence or noncoincidence of the infrared and Raman bands of HA-SRM. Hexagonal HA belongs to the space group P6_3 ; if, however, the OH^- ions are disregarded, the overall structure is $\text{P6}_3/\text{m}$. The lower P6_3 symmetry results from the position, heteronuclearity, and order of the OH^- ions. In fluoroapatite ($\text{P6}_3/\text{m}$ space group), the F ions are located along the c -axis on the mirror planes passing through the Ca_{II} triangles, whereas in hexagonal HA, the OH^- ions, with internuclear axes coincident with the c -axis, are displaced about 0.03 nm from the planes of the Ca_{II} triangles with protons pointing away from the Ca_{II} triangles [33]; thus, the mirror planes passing through the Ca_{II} triangles are lost and the P6_3 space group results. These minor structural differences cause considerable differences in the vibrational selection rules.

A comparison of the predicted and observed spectral data for the PO_4 modes in Table 4 shows a better fit with C_{6h} than with C_6 symmetry. Weights of 1, 2, 3, and 3 were applied to data for the ν_1 , ν_2 , ν_3 , and ν_4 PO_4

Table 4. Predicted number and coincidence or noncoincidence of infrared and Raman ν_1 , ν_2 , ν_3 , and ν_4 bands for PO_4 modes of hexagonal structures ($\text{P6}_3/\text{m}$, $\text{C}_{6\text{h}}$) and (P6_3 , C_6) of calcium hydroxyapatite^a and observed bands for HA-SRM

Hexagonal structure	Spectra	PO_4 modes			
		ν_1	ν_2	ν_3	ν_4
$\text{P6}_3/\text{m}$, $\text{C}_{6\text{h}}$	IR predicted	1nc	2nc	3nc	3nc
	R predicted	2nc	3nc	5nc	5nc
P6_3 , C_6	IR predicted	2c	4c	6c	6c
	R predicted	1nc, 2c	2nc, 4c	3nc, 6c	3nc, 6c
HA-SRM	IR observed	1c	2nc	7nc, 2c	4nc, 1c
	R observed	1c	2nc	3nc, 2c	3nc, 1c

^a Predicted from Ref. 21.
IR = infrared.
R = Raman.
c = coincident.
nc = noncoincident.

modes, respectively; these numerical weights correspond to the degeneracy of each mode. About 74 % of the total spectral data for the PO_4 modes (the total number of IR and Raman bands and the number of coincident/noncoincident bands) better fit with $\text{C}_{6\text{h}}$ symmetry. About 26 % of the total spectral data for the PO_4 modes (the total number of IR bands for the ν_3 and ν_4 PO_4 modes, the coincidence of the ν_1 IR and Raman PO_4 bands, and the coincidence/noncoincidence of the ν_3 Raman bands) better fit with C_6 symmetry. This better agreement with $\text{C}_{6\text{h}}$ symmetry is in accordance with previous conclusions based on fewer spectroscopic data [21,26] that also favored $\text{C}_{6\text{h}}$ symmetry ($\text{P6}_3/\text{m}$ space group) for hexagonal HA.

The number of observed IR ν_3 and ν_4 PO_4 bands is larger than predicted for $\text{C}_{6\text{h}}$ symmetry. This is believed

to derive from sources other than effects of lower P6_3 symmetry, and this will be considered in a separate paper [34].

3.6 X-Ray Diffraction Pattern

The XRD pattern of HA-SRM is shown in Fig. 7. The observed positions of diffraction lines (2θ and corresponding $d_{2\theta}$) and their relative intensities (I_{rel}) are listed in Table 5. These $d_{2\theta}$ and I_{rel} for HA-SRM are in full agreement with the corresponding values reported for hexagonal HA (JCPDS, Card No. 9-432) [35]. The additional weak lines of monoclinic HA that have relative intensities less than 1 % of the strongest hexagonal HA line were not observed at the intensity scale shown in Fig. 7. The additional XRD measurements, from

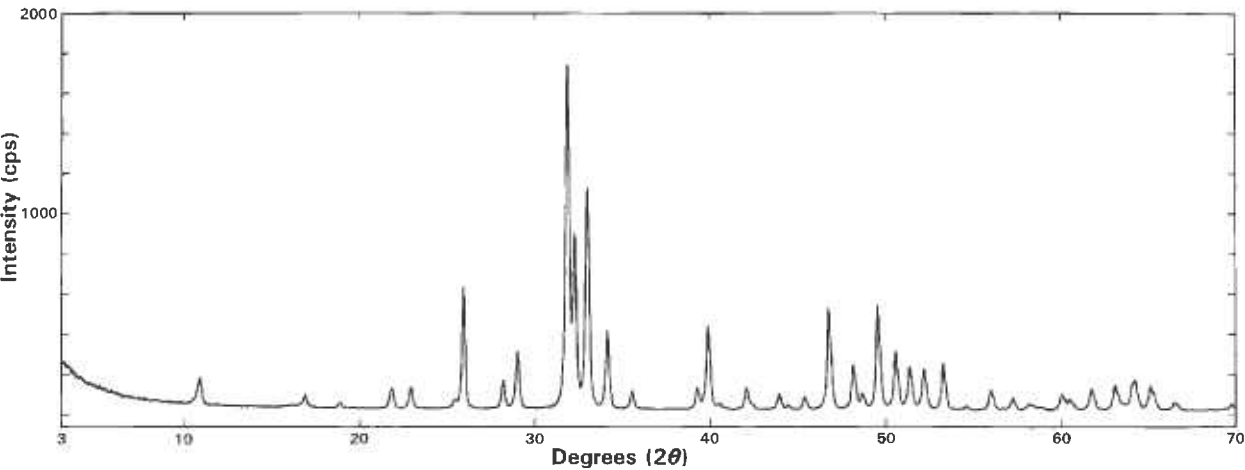


Fig. 7. X-ray diffraction pattern of HA-SRM in the 2θ range from 3° to 70° .

Table 5. 2θ -values and relative intensities (I_{rel}) observed from the XRD pattern of HA-SRM, d -values determined from 2θ -values ($d_{2\theta}$), d -values calculated from unit cell parameters (d_{calc}), and corresponding indices (hkl)

$2\theta(^{\circ})$	$d_{2\theta}(\text{nm})$	$d_{\text{calc}}(\text{nm})$	I_{rel}	hkl
10.85	0.815	0.816	8	100
16.87	0.525	0.526	3	101
18.84	0.471	0.471	2	110
21.75	0.408	0.408	6	200
22.84	0.389	0.389	6	111
25.35	0.351	0.351	2	201
25.86	0.344	0.344	36	002
28.11	0.317	0.317	8	102
28.92	0.308	0.308	16	210
31.77	0.281	0.281	100	211
32.18	0.278	0.278	47	112
32.90	0.272	0.272	65	300
34.04	0.263	0.263	22	202
35.44	0.253	0.253	5	301
39.18	0.2297	0.2297	6	212
39.793*	0.2263	0.2263	22	310
40.43	0.2229	0.2229	1	221
41.98	0.2150	0.2150	6	311
42.30	0.2135	0.2134	1	302
43.84	0.2063	0.2063	4	113
44.36	0.2040	0.2040	1	400
45.29	0.2000	0.2000	4	203
46.683*	0.1944	0.1944	28	222
48.068*	0.1891	0.1891	12	312
48.58	0.1872	0.1872	3	320
49.458*	0.1841	0.1841	30	213
50.474*	0.1807	0.1807	15	321
51.254*	0.1781	0.1781	11	410
52.061*	0.1755	0.1755	11	402
53.167*	0.1721	0.1721	14	004
54.43	0.1684	0.1684	1	104
55.85	0.1645	0.1645	6	322
57.11	0.1611	0.1611	4	313
58.03	0.1588	0.1588	2	501
58.28	0.1582	0.1582	2	412
58.74	0.1570	0.1570	1	330
59.93	0.1542	0.1542	4	420

* 2θ -values have expanded uncertainty (U) of $\pm 0.004^{\circ} 2\theta$ ($n = 4$).

which a mass fraction of about 25 % of monoclinic HA was determined in HA-SRM, will be reported in a separate paper [8].

3.7 Unit-Cell Parameters

The a and c unit-cell parameters for HA-SRM calculated from the eight selected diffraction lines (2θ -values marked with a in Table 5) are listed in Table 6. The complete set of d -values (d_{calc}) calculated from these unitcell parameters is listed in Table 5. These d_{calc} -values are in excellent agreement with $d_{2\theta}$ -values determined from the 2θ -values that were not used for unit-cell parameters calculation (2θ -values without asterisks in Table 5).

The a and c unit-cell parameters for HA-SRM determined in this paper are in very good agreement with the parameters determined for the same material by the Rietveld analyses [5,36], given in Table 6. The average values of these two independently determined unit-cell parameters for HA-SRM by the Rietveld analyses are: $a = 0.94235$ nm, and $c = 0.68852$ nm. As compared with these average unit-cell parameters, the values determined in this paper are 0.003 % larger in a , and 0.003 % larger in c than the corresponding average values. The values for similarly prepared hexagonal HA [9] determined by the Rietveld analyses [37] (Table 6, HA-McDowell) are 0.065 % smaller in a , and 0.001 % larger in c than the corresponding average values for HA-SRM determined by the Rietveld analyses.

3.8 Crystallinity

The mean angular widths at half-height (denoted as B and b) for the (200), (002), (102), (210), (310), and (004) diffraction lines of HA-SRM (B -values) and of hc-HA (b -values) and the calculated $1/\beta$ values are listed in Table 7. The $1/\beta$ values were determined in the next crystal directions: (i) along the a -axis perpendicular to b - c plane, $1/\beta(200) = 6.0 (^{\circ} 2\theta)^{-1} \pm 0.3 (^{\circ} 2\theta)^{-1}$, (ii) along the c -axis perpendicular to a - b plane, $1/\beta(002) = 8.4 (^{\circ} 2\theta)^{-1} \pm 0.2 (^{\circ} 2\theta)^{-1}$ and $1/\beta(004) = 7.1 (^{\circ} 2\theta)^{-1} \pm 0.3 (^{\circ} 2\theta)^{-1}$, (iii) perpendicular to c -axis, $1/\beta(210) = 5.8 (^{\circ} 2\theta)^{-1} \pm 0.2 (^{\circ} 2\theta)^{-1}$ and $1/\beta(310) = 5.5 (^{\circ} 2\theta)^{-1} \pm 0.2 (^{\circ} 2\theta)^{-1}$ and (iv) perpendicular to b -axis, $1/\beta(102) = 8.2 (^{\circ} 2\theta)^{-1} \pm 0.4 (^{\circ} 2\theta)^{-1}$. The bigger $1/\beta$ value denotes the larger crystal size and lattice perfection in corresponding crystal directions showing for HA-SRM the biggest $1/\beta$ values for size/strain in directions along c -axis and that perpendicular to b -axis, and the smallest $1/\beta$ values for size/strain in directions along a -axis and those perpendicular to c -axis. The $1/\beta(002)$ for HA-SRM is ≈ 10 % smaller and $1/\beta(310)$ for HA-SRM is ≈ 40 % larger than corresponding values for HA prepared by DCPA hydrolysis at pH ≈ 6.5 [38]. For HA-SRM the ratio of $1/\beta(002)$ and $1/\beta(310)$ values, $R(1/\beta) = [1/\beta(002)]/[1/\beta(310)] = \beta(310)/\beta(002)$, is 1.6 and for HA hydrolyzed from DCPA at pH ≈ 6.5 the ratio $R(1/\beta)$ is 2.5 [38]. These $R(1/\beta)$ -values can be correlated with the ratio of crystal height (longer dimension) and crystal width (shorter dimension) of these HA crystals determined microscopically. HA-SRM crystals for which $R(1/\beta) = 1.6$ have cylindrical shape with the height/diameter ratio of ≈ 2 (Fig. 2), and HA crystals hydrolyzed from DCPA, for which $R(1/\beta) = 2.5$, have plate-like shape with very large height/width ratio of ≈ 10 [38]. It indicates that the

Table 6. Unit-cell parameters for HA-SRM and similarly prepared HA by McDowell et al. [9]

Sample	<i>a</i> (nm)	<i>c</i> (nm)	XRD analysis	Reference
HA-SRM	0.94238 ± 0.00009 ^a	0.68854 ± 0.00006 ^a	Standard	This paper
HA-SRM	0.942253 ± 0.000013 ^a	0.688501 ± 0.000009 ^a	Rietveld	[5]
HA-SRM	0.94244 ± 0.00002 ^b	0.68854 ± 0.00002 ^b	Rietveld	[36]
HA-McDowell	0.94174 ± 0.00002 ^b	0.68853 ± 0.00002 ^b	Rietveld	[37]

^a Mean value ± expanded uncertainty (*U*).

^b Mean value ± standard deviation.

Table 7. The line width at half-height (*B*-value) for selected XRD lines of HA-SRM, the corresponding line width at half-height (*b*-value) of hc-HA^a, and calculated $1/\beta$ values

<i>hkl</i>	<i>B</i> (° 2θ)	<i>b</i> (° 2θ)	$1/\beta$ (° 2θ) ⁻¹
200	0.225 ± 0.007	0.150 ± 0.002	6.0 ± 0.3
002	0.188 ± 0.002	0.145 ± 0.002	8.4 ± 0.2
102	0.183 ± 0.005	0.136 ± 0.004	8.2 ± 0.4
210	0.218 ± 0.003	0.134 ± 0.003	5.8 ± 0.2
310	0.218 ± 0.004	0.120 ± 0.004	5.5 ± 0.2
004	0.181 ± 0.005	0.114 ± 0.003	7.1 ± 0.3

^a Highly crystalline HA prepared by solid state thermal reaction [17].

c-axis of these HA crystals is in the direction along the crystal height and the *a*-axis is in the direction along the crystal width.

3.9 Solubility

The solubility product of this HA-SRM was previously determined [4]. The saturated solutions with respect to HA-SRM were obtained by dissolution of HA-SRM crystals in aqueous solutions of phosphoric acid for 60 d at 37.0 °C ± 0.1 °C. The thermodynamic solubility product, K_{sp} , of HA-SRM defined as $K_{sp}(\text{HA}) = a^5(\text{Ca}^{2+}) a^3(\text{PO}_4^{3-}) a(\text{OH}^-)$, where *a* denotes ion activity, was calculated from measured equilibrium calcium and phosphate concentrations and pH values as input data. The mean value and standard uncertainty, u_i , of the twelve replicate determinations ($n = 12$) was $K_{sp}(\text{HA}) = (2.03 \pm 0.04) \times 10^{-59}$. The standard uncertainties, $u_i(y)$, derived from other sources were also determined. These other sources were uncertainties in measurements of Ca, P and pH, and uncertainties in dissociation constants of phosphoric acid (K_1 , K_2 , and K_3) and stability constant of calcium phosphate complexes used for K_{sp} calculation. The major contributions to the combined uncertainty, $u_c = 0.356 \times 10^{-59}$, were from pH measurements ($u_i = 0.196$) and the K_3 literature value ($u_i = 0.280$). The expanded uncertainty, $U = 2u_c$, was

0.71×10^{-59} ; thus, the thermodynamic $K_{sp}(\text{HA})$ at 37 °C, expressed as the mean ± *U*, was $(2.03 \pm 0.71) \times 10^{-59}$ and its $pK_{sp}(\text{HA})$ was 58.69 ± 0.15 . This $K_{sp}(\text{HA})$ value of $(2.03 \pm 0.71) \times 10^{-59}$ is in very good agreement with the literature value of $(2.36 \pm 0.28) \times 10^{-59}$ determined under similar conditions for similarly prepared HA [9].

4. Conclusions

The chemical and physical analyses of this HA-SRM are considered very reliable based on the consistency of the combined results. This HA-SRM has application as a standard of numerous well established chemical and physical properties to compare with and to establish the validity of equivalent analyses on natural and synthetic hydroxyapatites, the mineral phases in calcified tissues, and in testing and regulation.

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5. References

- [1] R. Z. LeGeros, Calcium Phosphates in Oral Biology, Karger, Basel (1991).
- [2] J. C. Elliott, Structure and Chemistry of the Apatites and Other Calcium Orthophosphates, Elsevier, Amsterdam (1994).
- [3] J. Arends, J. Christoffersen, M. R. Christoffersen, H. Eckert, B. O. Fowler, J. C. Heughebaert, G. H. Nancollas, J. P. Yesinowski, and S. J. Zawacki, A Calcium Hydroxyapatite Precipitated from an Aqueous Solution. An International Multimethod Analysis, *J. Crystal Growth* **84**, 515-532 (1987).

- [4] M. Markovic, B. O. Fowler, M. S. Tung, and E. S. Lagergren, Composition and Solubility Product of a Synthetic Calcium Hydroxyapatite. Chemical and Thermal Determination of Ca/P Ratio and Statistical Analysis of Chemical and Solubility Data, In: Mineral and Scale Formation, Proc. of ACS Symposium, Z. Amjad, ed., Plenum, New York (1995) pp. 271-282.
- [5] Certificate of Analysis, Standard Reference Material 2910, National Institute of Standards and Technology, Gaithersburg, MD, USA (1997).
- [6] B. O. Fowler, unpublished data.
- [7] J. C. Elliott and P. E. Mackie, Monoclinic Hydroxyapatite, In: Physico-chimie et Cristallographie des Apatites d'Interet Biologique—No 230, Centre National de la Recherche Scientifique (C.N.R.S.), Paris, 69-76 (1975).
- [8] M. Markovic, B. O. Fowler, and M. S. Tung, Preparation and Characterization of Monoclinic Calcium Hydroxyapatite, in preparation.
- [9] H. McDowell, T. M. Gregory, and W. E. Brown, Solubility of $\text{Ca}_5(\text{PO}_4)_3\text{OH}$ in the System $\text{Ca}(\text{OH})_2\text{-H}_3\text{PO}_4\text{-H}_2\text{O}$ at 5, 15, 25 and 37 °C, J. Res. Natl. Bur. Stand. (U.S.) **81A**, 273-281 (1977).
- [10] M. Markovic, B. O. Fowler, and W. E. Brown, Octacalcium Phosphate Carboxylates. 2. Characterization and Structural Considerations, Chem. Mater. **5**, 1406-1416 (1993).
- [11] A. Gee and V. R. Deitz, Determination of Phosphate by Differential Spectrophotometry, Anal. Chem. **25**, 1320-1324 (1953).
- [12] A. Gee and V. R. Deitz, Pyrophosphate Formation upon Ignition of Precipitated Basic Calcium Phosphates, J. Am. Chem. Soc. **77**, 2961-2965 (1955).
- [13] S. Brunauer, P. H. Emmett, and E. Teller, Adsorption of Gases in Multimolecular Layers, J. Am. Chem. Soc. **60**, 309-319 (1938).
- [14] R. N. Jones and A. Nadeau, Further Observations on the Use of Indene for the Wavenumber Calibration of Infrared Spectrometers, Spectrochim. Acta **20**, 1175-1183 (1964).
- [15] J. Loader, Chap. 3 in Basic Laser Raman Spectroscopy, Heyden, London (1970).
- [16] H. P. Klug and L. E. Alexander, X-ray Diffraction Procedures for Polycrystalline and Amorphous Materials, 2nd Ed., John Wiley and Sons, New York (1974) pp. 618-708.
- [17] B. O. Fowler, Infrared Studies of Apatites. II. Preparation of Normal and Isotopically Substituted Calcium, Strontium, and Barium Hydroxyapatites and Spectrastructurecomposition Correlations, Inorg. Chem. **13**, 207-214 (1974).
- [18] Guide to the Expression of Uncertainty in Measurement, ISBN 92-67-10188-9, 1st Ed., ISO, Switzerland (1993).
- [19] H. H. Ku, Notes on the Propagation of Error Formulas, J. Res. Natl. Bur. Stand. (U.S.) **70C**, 263-273 (1966).
- [20] H. M. Rootare and R. G. Craig, Vapor Phase Adsorption of Water on Hydroxyapatite, J. Dent. Res. **56**(12), 1437-1488 (1977).
- [21] B. O. Fowler, Infrared Studies of Apatites. I. Vibrational Assignments for Calcium, Strontium, and Barium Hydroxyapatites Utilizing Isotopic Substitution, Inorg. Chem. **13**, 194-207 (1974).
- [22] D. W. Holcomb and R. A. Young, Thermal Decomposition of Human Tooth Enamel, Calcif. Tissue Int. **31**, 189-201 (1980).
- [23] J. C. Elliott, The Crystallographic Structure of Dental Enamel and Related Apatites, PhD. Thesis, University of London (1974).
- [24] E. E. Berry, The Structure and Composition of Some Calcium-deficient Apatites, J. Inorg. Nucl. Chem. **29**, 317-327 (1967).
- [25] S. J. Joris and C. H. Amberg, The Nature of Deficiency in Nonstoichiometric Hydroxyapatites. II. Spectroscopic Studies of Calcium and Strontium Hydroxyapatites, J. Phys. Chem. **75**(20), 3172-3178 (1971).
- [26] K. C. Blakeslee and R. A. Condrate, Sr., Vibrational Spectra of Hydrothermally Prepared Hydroxyapatites, J. Amer. Ceram. Soc. **54**(11), 559-563 (1971).
- [27] D. C. O'Shea, M. L. Bartlett, and R. A. Young, Compositional Analysis of Apatites with Laser-Raman Spectroscopy: (OH, F, Cl) Apatites, Archs. Oral Biol. **19**, 995-1006 (1974).
- [28] B. O. Fowler, Raman and Infrared Vibrational Spectra of Apatites, Mineralized Tissue Research Communications **1**, Abstract No. 129 (1975). (Copy available from B. O. Fowler).
- [29] D. G. A. Nelson and B. E. Williamson, Low-temperature Laser Raman Spectroscopy of Synthetic Carbonated Apatites and Dental Enamel, Austral. J. Chem. **35**, 715-727 (1982).
- [30] B. O. Fowler, Polarized Raman Spectra of Apatites, J. Dent. Res. **56**, Abstract No. 68 (1977).
- [31] B. O. Fowler, I. Polarized Raman Spectra of Apatites. II. Raman Bands of Carbonate Ions in Human Tooth Enamel, Mineralized Tissue Research Communications **3**, Abstract No. 68 (1977). (Copy available from B. O. Fowler).
- [32] B. O. Fowler, M. Markovic, and W. E. Brown, Octacalcium Phosphate. 3. Infrared and Raman Vibrational Spectra, Chem. Mater. **5**, 1417-1423 (1993).
- [33] M. I. Kay, R. A. Young, and A. S. Posner, Crystal Structure of Hydroxyapatite, Nature **204**, 1050-1052 (1964).
- [34] B. O. Fowler, Second Derivative Vibrational Spectra of Fluoroapatite and Hexagonal and Monoclinic Hydroxyapatite, in preparation.
- [35] Powder Diffraction File: Inorganic Phases, Joint Committee on Powder Diffraction Standards, Swarthmore (1986) Card No. 9-432.
- [36] H. Morgan, R. M. Wilson, J. C. Elliott, S. E. P. Dowker, and P. Anderson, Preparation and Characterization of Monoclinic Hydroxyapatite and its Precipitated Carbonate Apatite Intermediate, Biomaterials **21**, 617-627 (2000).
- [37] R. A. Young and D. W. Holcomb, Variability of Hydroxyapatite Preparations, Calcif. Tissue Int. **34**, 517- 532 (1982).
- [38] K. Ishikawa and E. D. Eanes, The Hydrolysis of Anhydrous Dicalcium Phosphate into Hydroxyapatite, J. Dent. Res. **72**(2), 474-480 (1993).

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